

# COMPATIBILITY ASSESSMENT OF INDIGENOUS TRICHODERMA STRAINS WITH FUNGICIDE MOLECULES THROUGH IN VITRO ANALYSIS

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A study on compatibility of native strains of *Trichoderma* sp. with fungicides was ascertained at the Department of Plant Pathology, Dr Kalam Agricultural College, Kishanganj (under BAU, Sabour, Bhagalpur) during 2022-23. The compatibility studies revealed that among the eight fungicides tested (each @ 100, 250, and 500 ppm), carbendazim 50% WP, Propiconazole 25 EC, and Tebuconazole 25.9% EC were incompatible to all *Trichoderma* isolates. However, only three fungicides viz., Metalaxyl 4% + Mancozeb 64% WP, Mancozeb 75% WP and Copper oxychloride 50 % WP were found to be compatible and Azoxystrobin 250 EC, Azoxystrobin 18.2% W/W + Difenconazole 11.4% W/W was moderately compatible with all the test *Trichoderma* isolates. The compatible fungicides and doses of the test fungicides may be recommended for the farmers to manage plant diseases under integrated disease management practices.

Keywords : Biological control, Compatibility, Pesticides, Disease management.

# Introduction

*Trichoderma* is a genus of asexually reproducing saprophytic fungus that is found in almost all temperate to tropical soils, decaying plant tissues, and root ecosystems. *Trichoderma* spp. are effective biocontrol agents because they are potent antibiotic producers, rapid growers, prolific spore producers, and strong opportunistic invaders (Benitez *et al.* 2004). *Trichoderma* has a distinctive morphology that includes phialids, hyaline or green conidia carried in slimy heads, tufted or postulate conidiophores, and repeatedly branching conidiophores (Bisset 1984).

The genus *Trichoderma* is a member of the class Sordariomycetes, order Hypocreales, family Hypocreaceae, and phylum Ascomycetes. More than 200 years ago, Persoon (1794) in Germany published the first description of the genus *Trichoderma*. *Trichoderma* as bio-control agent was first reported by Weindling in 1932. Tulasne brothers described Hypocrea, a teleomorph of *Trichoderma*, in 1865. (Gams and Bissett, 2002). *Trichoderma* is an effective bio-control agent because of their high reproductive capacity, efficiency in nutrient uptake, ability to modify the rhizosphere, ability to be aggressive against plant pathogenic fungi, and effectiveness in promoting plant growth and defense mechanisms. This fungus, in addition to being used as a bio-control agent, serves as an excellent growth stimulant, solubilizes nutrients, produces numerous hydrolytic enzymes, and induces resistance in plants to various abiotic and biotic stresses. (Harman, 2011).

Commercial preparations of *Trichoderma* species have been utilised for biological management of fungal-induced plant diseases. *Trichoderma harzianum* is the active component in the product TRICHODEX, which is used to treat apple postharvest rot, and it is coupled with *T. polysporum* in the product BINAB-T, which is used to treat wound decay and wood rot. 620

(Ricard, 1981) The most widely utilised species in biological control are *T. harzianum and T. virens*. (Papavizas, 1985). *T. reesei's* capacity to break down cellulosic materials by producing cellulase enzymes has led to commercial use (Kubicek *et al.*, 1996). Because of their wide metabolic capabilities and intense competitive character, they were excellent colonisers of their environments. (Gams and Bissett, 2002).

*Trichoderma* biocontrol mechanism involves coiling around the host, the production of appressoria and the disintegration of the host cell wall, antibiosis, and competition for resources such as space and nutrients. (Tiwari *et al.*, 2021)

T. virens, T. atroviride, and T. reesei were the first species in the genus to have their genome sequenced, allowing for more in-depth research of the genus Trichoderma. (Guzman et al.. 2023). Trichoderma secondary metabolites can activate disease-fighting systems inside plants and protect them against infections. Exogenous indicators that might be utilised to identify and monitor specific Trichoderma isolates in agro-ecosystems include glucuronidase (GUS), green fluorescent protein (GFP), hygromycin B phosphotransferase (HygB), and generating genes. (Manzar et al., 2022).

# **Materials and Methods**

Soil samples were collected from the rhizosphere soil of Zone II of Bihar and were sieved, shade dried and used for isolation of *Trichoderma using* serial dilution and pour plate technique. Further, sub-cultured on the PDA medium to get the pure cultures using hyphal tip technique (Tuite, 1969) and their pure cultures were preserved in the refrigerator and used for further experiments.

# Compatibility of Trichoderma spp. with fungicides

In the present study eight fungicides, were selected for testing the compatibility with fungal biocontrol agents at various concentrations, by applying Poisoned food technique (Nene and Thapliayal, 1993), and using PDA as basal culture medium.

Required quantity of fungicides for each chemical was calculated using the formula,  $N_1V_1 = N_2V_2$ . In order to achieve the correct concentrations, the necessary quantity of the test fungicides was mixed with autoclaved and chilled (to room temperature) PDA medium in conical flask. Poisoned PDA with the test fungicides was put aseptically into petri plates (20 ml per plate). Three replications were maintained for each of the test fungicides and its test concentrations using CRD (Completely Randomized Design). After solidification of PDA medium, each plate was individually and aseptically inoculated with 5.0 mm mycelial disc taken from an actively growing mycelium of seven-day-old pure culture of *Trichoderma* sp. such as TR 1, TR 2, TR 3, TR 4, TR 5, TR 6 and incubated for four days at a temperature of 28°C. Petri plates poured with PDA (without any fungicides) and inoculated separately with TR 1, TR 2, TR 3, TR 4, TR 5, TR 6 and were maintained as untreated control.

#### Assessment of growth

Colony growth of *Trichoderma* isolates was assessed by measuring the diameter (mm) of mycelial growth using the measuring scale after four days of incubation. Per cent inhibition of colony growth was calculated by using the formula given by Vincent (1947).

I = [(C-T) / C] \* 100

Were,

I = Percent growth inhibition.

C = Colony diameter in control (mm).

T = Colony diameter in treatment (mm).

## Statistical analysis

The mycelial growth diameter and percentage inhibition of *Trichoderma* isolates were analyzed using OPSTAT software, which conducted a statistical examination of factorial CRD data.

**Table 1:** Treatment details of compatibility study of

 *Trichoderma* sp. with fungicides

Treatment detail										
Name of Fungicide	Concentration (ppm)									
Mancozeb 75% WP	100, 250, 500									
Propiconazole 25 EC	100, 250, 500									
Tebuconazole 25.9% EC	100, 250, 500									
Copper oxychloride 50% WP	100, 250, 500									
Carbendazim 50% WP	100, 250, 500									
Azoxystrobin 18.2% W/W + Difenconazole 11.4% W/W	100, 250, 500									
Azoxystrobin 250 EC	100, 250, 500									
Metalaxyl 4% + Mancozeb 64% WP Control (untreated)	100, 250, 500									

#### **Results and Discussion**

The result revealed that among the eight test fungicides, four systemic fungicides *viz.*, Carbendazim 50% WP, Propiconazole 25 EC, Tebuconazole 25.9% EC, Azoxystrobin 250 EC and two contact fungicides (Mancozeb 75% WP, Copper oxychloride 50% WP) and two combi- products (Azoxystrobin 18.2% W/W + Difenconazole 11.4% W/W, Metalaxyl 4% + Mancozeb 64% WP) at three concentrations viz., 100, 250, 500 ppm evaluated against *Trichoderma* isolates under *in vitro* conditions indicated that lower concentration (100-250ppm) of fungicides were found to be highly compatible with *Trichoderma* sp. as compared to higher concentration (500 ppm). Among, eight fungicides tested Carbendazim 50% WP, Propiconazole 25 EC, and Tebuconazole 25.9% EC were incompatible with all the *Trichoderma* isolates at all concentrations (100, 250 and 500 ppm) by inhibiting cent per cent mycelial growth of all the isolates. The data are presented in the Table 2, 3, and 4.

## At 100 ppm

Metalaxyl 4% + Mancozeb 64% WP, Mancozeb 75% WP and Copper oxychloride 50% WP were found to be compatible with all the isolates (Tr 1, Tr 2, Tr 3, Tr 4, Tr 5 and Tr 6) showing 0% inhibition and growth of colony was 90 mm. In Azoxystrobin 250 EC treated plates, isolate Tr 2 showed maximum inhibition 11.11% followed by Tr 6 (10.74%), Tr 1 (5.93%), Tr 4 (4.07%) and Tr 3 (3.93%) at 100 ppm concentration.

In case of Azoxystrobin 18.2% W/W + Difenconazole 11.4% W/W Isolate Tr 4 showed maximum inhibition (73.70%) followed by Tr 2 (25.56%), Tr 5 (22.22%), Tr 6 (13.70) and Tr 3 (13.70) at 100 ppm. Carbendazim 50% WP, Propiconazole 25 EC, and Tebuconazole 25.9% EC were incompatible with all the *Trichoderma* isolates at 100 ppm (Table 2, Fig 1).

#### At 250 ppm

Most of the tested Trichoderma isolates (Tr 1, Tr 2, Tr 3 and Tr 4) were found compatible with (Metalaxyl 4% + Mancozeb 64% WP) and Mancozeb 75% WP. Tr 2 and Tr 3 were found compatible with copper oxychloride 50% WP. While all the Trichoderma isolates Tr 1 to Tr 6 were completely inhibited by Carbendazim 50% WP, Propiconazole 25 EC, and Tebuconazole 25.9% EC indicating noncompatibility with these fungicides at 250 ppm. All the tested Trichoderma isolates showed partial compatibility with Azoxystrobin 18.2% W/W + Difenconazole 11.4% W/W (Table 3, Fig 2).

## At 500 ppm

Trichoderma isolates Tr 1, Tr 2, Tr 3, Tr 5 and Tr 6 were found to be compatible with Metalaxyl 4% + Mancozeb 64% WP with less than 10% inhibition of colony growth. Tr 1 and Tr 4 were found to be compatible with Mancozeb 75% WP with less than 20% inhibition. Isolates Tr 1, Tr 2, Tr 5 and Tr 6 were performed better than less than 20% inhibition with Copper oxychloride 50% WP. With Azoxystrobin 250 EC Tr 3 and Tr 6 found compatible with less than 20% inhibition. All the Trichoderma isolates were found non-compatible with Carbendazim 50% WP. Propiconazole 25 EC, and Tebuconazole 25.9% EC with Cent percent inhibition. Data is shown in (Table 4, Fig 3).

Table 2: Compatibility of *Trichoderma* isolates with different fungicides at 100 ppm

Trichoderm a	Metalaxyl 4% + Mancozeb 64 % WP		Mancozeb 75 % WP		Copper oxychloride 50 % WP		Azoxystrobin 250 EC		Azoxystrobin 18.2 % W/W+ Difenconazole 11.4 % W/W				Tebuconazole 25.9 % EC		Propiconazole 25 EC	
Isolates	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)
Tr 1	90.00	0.00	90.00	0.00	90.00	0.00	84.67	5.93	90.00	0.00	0.00	100.00	0.00	100.00	0.00	100.00
Tr 2	90.00	0.00	90.00	0.00	90.00	0.00	80.00	11.11	67.00	25.56	0.00	100.00	0.00	100.00	0.00	100.00
Tr 3	90.00	0.00	90.00	0.00	90.00	0.00	86.47	3.93	86.67	3.70	0.00	100.00	0.00	100.00	0.00	100.00
Tr 4	90.00	0.00	90.00	0.00	90.00	0.00	86.33	4.07	23.67	73.70	0.00	100.00	0.00	100.00	0.00	100.00
Tr 5	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	70.00	22.22	0.00	100.00	0.00	100.00	0.00	100.00
Tr 6	90.00	0.00	90.00	0.00	83.33	7.41	80.33	10.74	77.67	13.70	0.00	100.00	0.00	100.00	0.00	100.00
Control	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.0	0.00	90.00	0.00
CD @ 1 %	N/S		N/S		2.47		4.571		3.26		0.782		1.156		1.744	
C.V.	Nil		Nil		1.545		3.003		2.62		3.439		5.086		7.673	

\*Average of three replications

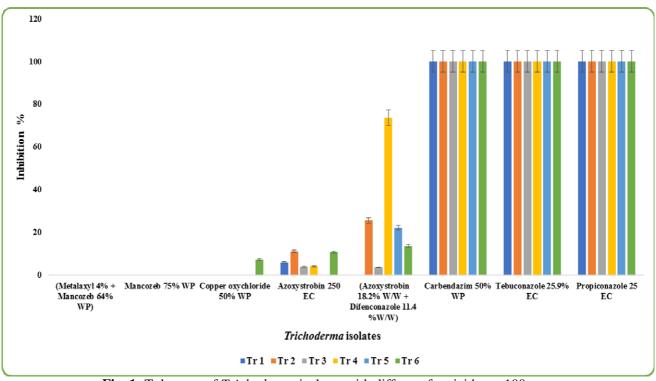


Fig. 1: Tolerance of Trichoderma isolates with different fungicides at 100 ppm

Trichoderma Isolates	Metalaxyl 4%+Mancozeb 64 % WP		Mancozeb 75% WP		Copper oxychloride 50% WP		Azoxystrobin 250 EC		Azoxystrobin 18.2 % W/W + Difenconazole 11.4 % W/W		- Carbendazim		Tebuconazole 25.9 % EC		Propiconazole 25 EC	
	G (mm)*	I(%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I(%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)
Tr 1	90.00	0.00	90.00	0.00	83.00	7.78	62.33	30.74	27.33	69.63	0.00	100.00	0.00	100.00	0.00	100.00
Tr 2	90.00	0.00	90.00	0.00	90.00	0.00	77.67	13.70	25.67	71.48	0.00	100.00	0.00	100.00	0.00	100.00
Tr 3	90.00	0.00	90.00	0.00	90.00	0.00	83.33	7.41	45.00	50.00	0.00	100.00	0.00	100.00	0.00	100.00
Tr 4	90.00	0.00	90.00	0.00	35.33	60.74	44.67	50.37	23.00	74.44	0.00	100.00	0.00	100.00	0.00	100.00
Tr 5	87.17	3.70	86.67	3.70	88.50	1.67	80.67	10.37	16.67	81.48	0.00	100.00	0.00	100.00	0.00	100.00
Tr 6	83.33	7.04	83.67	7.04	80.33	10.74	79.33	11.85	32.67	63.70	0.00	100.00	0.00	100.00	0.00	100.00
Control	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00
C.D@1%	2.705		3.498		3.321		4.312		1.427		N/A		N/A		N/A	
C.V.	1.701		2.2		2.371		3.361		2.795		2.7		2.9		2.3	

Table 3: Compatibility of Trichoderma isolates with different fungicides at 250 ppm

\*Average of three replications

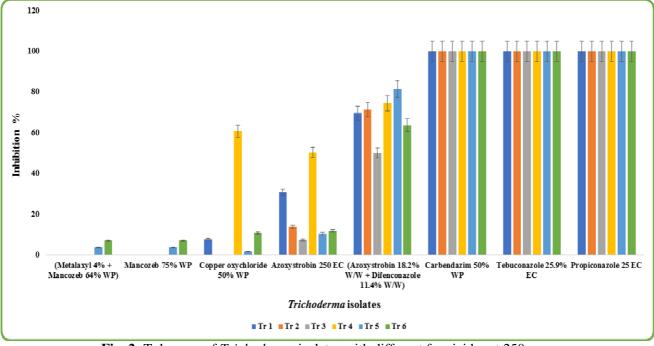


Fig. 2: Tolerance of Trichoderma isolates with different fungicides at 250 ppm

Trichoderma Isolates	Metalaxyl 4% + Mancozeb 64 % WP		Mancozeb 75 % WP		Copper oxychloride 50 % WP		Azoxystrobin 250 EC		Azoxystrobin 18.2 % W/W + Difenconazole 11.4 % W/W		- Carbendazim		Tebuconazole 25.9 % EC		Propiconazole 25 EC	
	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)	G (mm)*	I (%)
Tr 1	90.00	0.00	74.67	17.04	75.33	16.30	59.33	34.07	23.00	74.44	0.00	100.00	0.00	100.00	0.00	100.00
Tr 2	86.67	3.70	29.00	67.78	76.00	15.56	66.00	26.67	14.00	88.44	0.00	100.00	0.00	100.00	0.00	100.00
Tr 3	88.40	1.78	46.00	48.89	90.00	0.00	78.33	12.96	12.00	86.67	0.00	100.00	0.00	100.00	0.00	100.00
Tr 4	56.67	37.04	82.33	8.56	30.33	66.30	20.90	76.78	12.17	86.48	0.00	100.00	0.00	100.00	0.00	100.00
Tr 5	84.00	6.67	38.33	57.41	88.00	2.22	70.33	21.85	6.93	92.30	0.00	100.00	0.00	100.00	0.00	100.00
Tr 6	80.67	10.37	40.00	55.56	79.67	11.48	76.67	14.81	18.00	80.00	0.00	100.00	0.00	100.00	0.00	100.00
Control	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00	90.00	0.00
C.D@1%	4.713		1.573		2.87		2.916		0.846		1.14		1.323		0.962	
C.V.	3.232		1.691		2.179		2.618		3.276		5.03		5.821		4.233	

Table 4: Compatibility of Trichoderma isolates with different fungicides at 500 ppm

\*Average of three replications

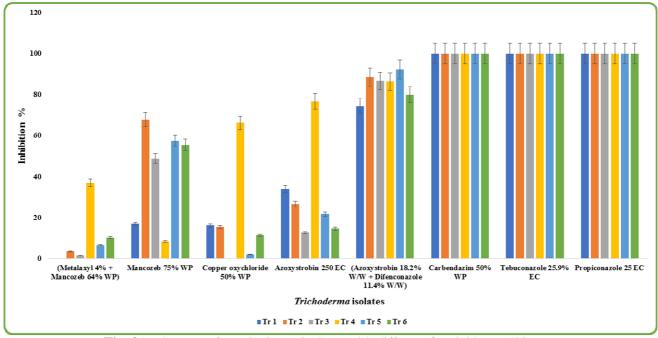


Fig. 3: Tolerance of Trichoderma isolates with different fungicides at 500 ppm

The results were closely supported with the findings of Tomer et al. (2018) who conducted an experiment to test the compatibility of four fungicides viz. Mancozeb, Thiram, Carboxin and Propiconazole with Trichoderma harzianum at 25, 50, 75 and 100 ppm and found that Mancozeb was highly compatible with all the four concentrations. Thiram was less compatible than Mancozeb and Carboxin and Propiconazole was toxic and incompatible with Trichoderma harzianum. Mishra et al. (2019)evaluated nine fungicides for their compatibility with bioagent Trichoderma viride. He observed that Mancozeb showed least inhibition of 42.96% at 200 ppm. Five other fungicides namely Azoxystrobin, Tebuconazole, Hexaconazole, Carbendazim, Propiconazole completely inhibited the growth and hence not compatible with Trichoderma viride. Kumar et al. (2019) revealed that T. viride was not compatible with Carbendazim 50%WP, Propiconazole 25%EC and Hexaconazole 5%EC even at 50 ppm concentration and 100 per cent inhibition in growth of T. viride was observed at 50 ppm and above concentrations. Thus the findings of current study is supported by the findings of Kumar et al. . Maheshwary et al. (2020) tested the compatibility of Trichoderma asperellum with different fungicides where they found that Trichoderma asperellum was most compatible with Copper hydroxide, Copper oxychloride, Metalaxyl, Mancozeb and least compatible with Captan. Whereas it was incompatible with Carbendazim, Tebuconazole and Propiconazole. Among combination fungicides Metalaxyl-M + Mancozeb was extremely compatible

with Trichoderma asperellum. These findings are in accordance with current findings with respect to compatibility of Metalaxyl + Mancozeb, Mancozeb, Copper oxychloride and incompatibility of Carbendazim, Tebuconazole and Propiconazole with Trichoderma isolates. Poudel et al. (2023) reported that 100 ppm of Copper oxychloride and 100,200 and 300 ppm of Mancozeb and Metalaxyl + Mancozeb was found to be compatible with T. harzianum and highest inhibition (100%) was observed in Carbendazim + Mancozeb, Carbendazim and Hexaconazole even at lower concentration of 100 ppm. These findings also support the findings of present studies.

#### Conclusion

Among eight tested fungicides (each @ 100, 250, and 500 ppm), Carbendazim 50% WP, Propiconazole 25 EC, and Tebuconazole 25.9% EC showed 100 per cent inhibition of mycelial growth indicating the incompatibility of these fungicides with all tested Trichoderma isolates. However, only three fungicides viz., Metalaxyl 4% + Mancozeb 64% WP, Mancozeb 75% WP and Copper oxychloride were found to be compatible with all the test Trichoderma isolates at lower concentrations of 100 and 250 ppm. While Azoxystrobin 250 EC, Azoxystrobin 18.2% W/W + Difenconazole 11.4% W/W showed moderately compatibility with all test Trichoderma isolates at all concentrations. The compatible fungicides may further screen at field conditions and can be used for sustainable and eco-friendly management of crop diseases under integrated disease management practices in order to reduce the use of harmful and higher doses of inorganic fungicides.

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#### **Competing Interests**

Authors have declared that no competing interests exist.

#### References

- Benitez, T., Rincon, A.M., Limon, M.C. and Codon, A.C. (2004) Biocontrol mechanisms of *Trichoderma* strains. *International Microbiology*, 7: 249- 260.
- Bissett, J. (1984). A revision of the genus *Trichoderma:* 1. Section Longibrachiatum, new section. *Canadian Journal* of Botany, 62, 924–931.
- Gams, W. and Bissett, J. (2002). Morphology and identification of *Trichoderma*. In Kubicek CP Harman GE. *Trichoderma* and *Gliocladium*: Basic biology, taxonomy and genetics. Taylor & Francis Ltd, pp. 3-31.
- Guzman, P., Kumar, A., de Los Santos-Villalobos, S., Parra-Cota, F.I., Orozco-Mosqueda, M.D.C., Fadiji, A.E., Hyder, S., Babalola, O.O., Santoyo, G. (2023). *Trichoderma* Species: Our Best Fungal Allies in the Biocontrol of Plant Diseases-A Review. Plants, 12(3), 432.
- Harman, G.E. (2011). *Trichoderma* not just for biocontrol anymore. *Phytoparasitica*, 39(2), 103-108.
- Kubicek, C.P., Bolzlbauer, U.M., Kovacs, W., Mach, R.L., Kuhls, K., Lieckfeldt, E., Borner, T., Samuels, G.J. (1996). Cellulase formation by species of *Trichoderma* sect. Longibrachiatum and of *Hypocrea* spp. with anamorphs referable to *Trichoderma* sect. Longibrachiatum. *Fungal Genetic and Biology*, 20, 105– 114.
- Kumar, A., Bansal, R.D. and Chelak, Y.K. (2019). Compatibility of *Trichoderma viride* with Fungicides for Plant Disease Management, *International Journal of Pure* and Applied Bioscience, 7(3), 44-51.
- Maheshwary, N., Gangadhara Naik, B., Amoghavarsha, C.M., Naik, S.K. and Nandish, M. (2020). "Compatibility of

*Trichoderma asperellum* with fungicides. *Pharma Innovation* 9, 136-140.

- Manzar, N., Kashyap, A.S., Goutam, R.S., Rajawat, M.V.S., Sharma, P.K., Sharma, S.K., Singh, H.V. (2022). *Trichoderma*: Advent of Versatile Biocontrol Agent, Its Secrets and Insights into Mechanism of Biocontrol Potential. *Sustainability*, 14(19), 12786.
- Mishra, S., Mishra, P., Singh, R., Singh, G. and Sachan, S.K. (2019). Compatibility of Different Systemic and Non Systemic Fungicides with *Trichoderma viride*. *International Journal of Current Microbiology and Applied Science*, 8(1), 1005-1010.
- Nene, Y.L. and Thapliyal, P.N. (1993). Fungicides in plant disease control. International Science Publisher. Oxford and IDH Publication Company, pp. 507.
- Papavizas, G.C. (1985). Trichoderma and Gliocladium: Biology and potential for biological control, Annual Review of Phytoapthology, 23, 23-54.
- Persoon, C.H. (1794). Production of non- volatile antibiotics. *Romers Neues Mag Bot*, 1: 81-128.
- Poudel, S., Pun, L.B., Paudel, R. and Thapa, S. (2023). *In-vitro* Compatibility Assessment of *Trichoderma harzianum* with Chemical Fungicides and Botanical Extracts. *Journal* of the Institute of Agriculture and Animal Science, 37, 121-131.
- Ricard, J.L. (1981). Commercialization of a *Trichoderma*-based mycofungicide: some problems and solutions, Biocontrol News and Information, 2(2), 95-98.
- Tiwari, R., Shukla, S.K., Jaiswal, V.P., Sharma, L., Joshi, D., Chandra, K., Asha Gaur, Srivastava Abhay, Kumar Rajesh, Tiwari, R.K. (2021). Bio-control potential of *Trichoderma* spp., against *Fusarium* spp., the incitants of Pokkah boeng disease of sugarcane under *in-vitro* conditions. *Indian Phytopathology*, 74, 691-701.
- Tomer, A., Singh, R. and Prasad, D. (2018). Compatibility *Trichoderma harzianum* with systemic and two non systemic fungicides of *in vitro*. *Asian Journal of Crop Science*, 10: 174-179.
- Tuite, J. (1969). Plant pathological methods, fungus and bacteria. Vol. I Burgess Publishing CO, Minneapolis, USA. P. 238.
- Vincent, J.M. (1947). Distortion of fungal hyphae in the presence of certain inhibitors. *Nature*, 150: 850.
- Weindling, R. (1932). Trichoderma lignorum as a parasite of other soil fungi. Phytopathology, 22: 837-845.